

MYELOID-DERIVED GROWTH FACTOR (MYDGF) (MOUSE) ULTRASENSITIVE ELISA KIT

FOR THE QUANTITATIVE DETERMINATION
OF MOUSE MYGDF CONCENTRATIONS
SERUM, AND PLASMA



THIS ELISA KIT IS ONE TIME USE
ONLY.

FOR RESEARCH USE ONLY. NOT FOR USE IN
DIAGNOSTIC PROCEDURES.

PURCHASE INFORMATION:

ELISA NAME	MYGDF (MOUSE) ULTRASENSITIVE ELISA KIT
Catalog No.	SK00579-03
Lot No.	
Formulation	96 T
Standard Range	3.9 - 250 pg/mL
Sensitivity	3 - 5 pg/mL
Sample Volume	75 µl
Sample Type	Serum, EDTA Plasma
Dilution Factor	Optimal dilutions should be determined by each laboratory for each application
Specificity	Human
Calibration	Mouse MYGDF Rec
Intra-assay Precision	4-6%
Inter-assay Precision	10-12%
Storage	2 - 8° C

Order Contact:
AVISCIERA BIOSCIENCE, INC
2348 Walsh Ave, Suite C
Santa Clara, CA 95051
USA
Tel: (408) 982 0300
Email: Sales@AvisceraBioscience.com
Website: www.AvisceraBioscience.com

INTRODUCTION

Mouse Myeloid-Derived Growth Factor (MYDGF) / SF20 Ultrasensitive immunoassay Kit is a solid phase ELISA designed to measure mouse MYGDF serum, and plasma. It contains recombinant mouse MYGDF and antibodies raised against this protein. It has been shown to accurately quantify recombinant mouse MYGDF and relative mass values for natural mouse MYGDF.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for mouse MYGDF has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any mouse MYGDF present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated antibody specific for mouse MYGDF is added to the wells. Following a wash to remove any unbound antibody reagent, Streptavidin HRP is added to the wells. After a final wash cycle, Chemifluorescent Substrate working solution was added for 30 minutes incubation before adding the Stop Solution. The fluorescent signal was measured using a Fluorescent microplate reader with excitation/emission at 325/420nm.

LIMITATIONS OF THE PROCEDURE

- _ FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- _ The kit should not be used beyond the expiration date on the kit label.
- _ Do not mix or substitute reagents with those from other lots or sources.
- _ It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.
- _ If samples generate values higher than the highest standard, dilute the samples with the appropriate Dilution Buffer and repeat the assay.
- _ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- _ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors have been tested in the immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
Mouse MYGDF Microplate - 96 well polystyrene black microplate (12 strips of 8 wells) coated with an antibody against mouse MYGDF.	579-03-01	1 plate
Mouse MYGDF Standard – refer to package label of recombinant mouse MYGDF in a buffered protein base with preservative; lyophilized.	579-03-02	1 vial
Detection Antibody Concentrate – refer to package label of vial, 100-fold concentrated of a biotinylated antibody against mouse MYGDF with preservative; lyophilized.	579-03-03	1 vial
Positive Control – one vial of recombinant mouse MYGDF; lyophilized.	579-03-04	1 vial
Streptavidin-HRP Conjugate – refer to package label of vial, 100-fold concentrated solution of Streptavidin-HRP conjugate.	SAHRP	1 vial
Dilution Buffer- 60 mL of buffered protein based solution with preservative.	DB01	1 bottle
Wash Buffer - 50 mL of 10-fold concentrated buffer with preservative.	WB01	1 bottle
Chemifluorescent Solution A – 8 mL of Chemifluorescent substrate solution.	CFS02A	1 bottle
Chemifluorescent Solution B – 1 mL of Chemifluorescent substrate solution.	CFS02B	1 bottle
STOP Solution C – 8 mL/vial	CFS02-STOP	1 vial
Plate Sealer	EAPS	1
Plastic Pouch	P01	1

STORAGE

Unopened Kit: Store at 2 - 8° C for up to 6 months. For longer storage, unopened Standard, Positive Control, and Detection Antibody Concentrate should be stored at -20° C or -70° C. Do not use kit past expiration date.

THIS ELISA IS ONE TIME USE ONLY.

Microplate Wells: Return unused wells to the plastic pouch with the desiccant pack. Microplate may be stored for up to 6 month at 2 - 8° C.

OTHER SUPPLIES REQUIRED

- Fluorescent microplate reader with excitation/emission at 325/420nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

SAMPLE COLLECTION AND STORAGE

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20° C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20° C. Avoid repeated freeze-thaw cycles.

Optional: Use Aprotinin (enzyme inhibitor) for ALL sample collection to prevent sample degradation. 0.5 TIU per ml of sample solution.

SAMPLE PREPARATION

Optimal dilutions should be determined by each laboratory for each application. Use polypropylene test tubes.

REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of 1x Wash Buffer.

Human MYGDF Standard - Refer to vial label for reconstitution volume. Reconstitute the human MYGDF standard with refer to lot specific package label of Dilution Buffer. This reconstitution produces a stock solution of refer to lot specific package label

pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 µL of Dilution Buffer into tubes #2 to #6. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 250 pg/mL standard serves as the high standard. The appropriate Dilution Buffer serves as the zero standard (0 pg/mL).

TUBE	STANDARD	DILUTION BUFFER	CONCENTRATION
Stock	Powder	Refer to lot specific	Refer to lot specific
# 1	Refer to lot specific protocol		250 pg/ml
# 2	250 µl of 1	250 µl	125 pg/ml
# 3	250 µl of 2	250 µl	62.5 pg/ml
# 4	250µl of 3	250 µl	31.25 pg/ml
# 5	250 µl of 4	250 µl	15.6 pg/ml
# 6	250 µl of 5	250 µl	7.8 pg/ml
# 7	250 µl of 6	250 µl	3.9 pg/ml

Positive Control - Reconstitute the Positive Control stock with 1 mL of Dilution Buffer. Allow the standard to sit for a minimum of 15 minutes with gentle agitation. Positive control should be prepared and used immediately.

Detection Antibody - Reconstitute the Detection Antibody Concentrate with refer to package label of Dilution Buffer to produce a 10-fold concentrated stock solution.

Streptavidin HRP Conjugate - Pipette 7.92 mL of Dilution Buffer into a 15 ml centrifuge tube and transfer 80 µl of 100-fold concentrated stock solution to prepare working solution. **Note:** 1x working solution of Streptavidin HRP conjugate should be used immediately.

Chemifluorescent Mix Solution – Mix 7.2ml (9 parts) of Solution A and 0.8 mL (1 part) of Solution B. **Note:** Bring all substrate solutions to room temperature before use.

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards, positive control and samples be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the plastic pouch with the desiccant pack.
3. Add 75 µL of Dilution Buffer to Blank wells.
4. Add 75 µL of Standard solutions in reverse order of serial dilution, sample, or positive control per well. Cover with plate sealer. Incubate for 2 hours on microplate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash with **1x Wash Buffer**, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (300 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 75 µL of Detection Antibody working solution to each well. Cover with plate sealer. Incubate for 2 hours on microplate shaker at room temperature.
7. Repeat the aspiration/wash as in step 5.
8. Add 75 µL of **Streptavidin-HRP Conjugate** working solution to each well. Incubate for 60 minutes on microplate shaker at room temperature. **Protect from light.**
9. Repeat the aspiration/wash as in step 5.
10. Add 75 µL of Chemifluorescent Mix Solution to each well. Incubate for 30 minutes on microplate shaker at room temperature. **Protect from light.**
11. Add 75 µL of Stop Solution C to each well. Shake the plate to ensure thorough mixing.
12. The fluorescent signal was measured using a fluorescent microplate reader with excitation/emission at 325/420nm.

CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control and sample and subtract the average zero standard optical density. Create a standard curve by reducing the data using the reader’s software. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the soluble neprilysin

concentrations versus the log of the absorbance and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

CALIBRATION

This immunoassay is calibrated against a highly purified recombinant mouse MYGDF.

SENSITIVITY

The minimum detectable dose (MDD) of mouse MYGDF was 5 pg/mL.

TYPICAL DATA









This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

STANDARD (PG/ML)	RFU
3.9	112249
7.8	290121
15.6	632200
31.25	1224384
62.5	2247936
125	4228720
250	8999442

SPECIFICITY

PROTEINS	CROSS-REACTIVITY (%)
Human MYGDF	100
Human IL-33	0
Human IL-6	0
Human FSTL1	0

SUMMARY OF ASSAY PROCEDURE

PREPARE REAGENTS, SAMPLES AND STANDARDS

Add 75 µl of standard, samples, positive control to each well. Incubate 2 hours on the plate shaker at RT.

Aspirate and wash 4 times with 1x Wash Buffer.

Add 75 µl Detection Antibody working solution to each well. Incubate 2 hours on the plate shaker at RT.

Aspirate and wash 4 times with 1x Wash Buffer.

Add 75 µl Streptavidin HRP conjugate working solution to each well. Incubate 60 min on the plate shaker at RT. Protect from light.

Aspirate and wash 4 times with 1x Wash Buffer.

Add 75 µl Chemifluorescent Mix Solution to each well. Incubate 30 min on the plate shaker at RT. Protect from light.

Add 75 µl Stop Solution to each well. Read Fluorescent signal with Ex 325nm, Em 420nm