

HUMAN PROINSULIN ELISA KIT

FOR THE QUANTITATIVE DETERMINATION OF
HUMAN PROINSULIN CONCENTRATIONS IN
CELL CULTURE SUPERNATES, SERUM AND
PLASMA



FOR RESEARCH USE ONLY. NOT FOR USE IN
DIAGNOSTIC PROCEDURES.

PURCHASE INFORMATION:

ELISA NAME	HUMAN PROINSULIN ELISA
Catalog No.	SK00095-01
Lot No.	
Formulation	96 T
Standard range	0.234 – 15 ng/mL
Sensitivity	80 pg/mL
Sample Volume	100 µL
Sample Type	Serum, EDTA Plasma, Cell Culture Supernates
Dilution Factor	<i>Optimal dilutions should be determined by each laboratory for each application</i>
Specificity	Human proinsulin
Intra-assay Precision	4 - 6%
Inter-assay Precision	8 - 10%
Storage	2 – 8 °C

ORDER CONTACT:

AVISCERA BIOSCIENCE, INC.

2348 Walsh Ave., Suite C

Santa Clara, CA 95051

USA

Tel: (408) 982 0300

Fax: (408) 982 0301

Email: Sales@AvisceraBioscience.com

Info@AvisceraBioscience.com

www.AvisceraBioscience.com

INTRODUCTION

Human PROINSULIN immunoassay is a solid phase ELISA designed to measure human PROINSULIN in cell culture supernates, serum and plasma. It contains recombinant human PROINSULIN and antibodies raised against this protein. It has been shown to accurately quantify recombinant human PROINSULIN. Results obtained with naturally occurring PROINSULIN samples showed linear curves that were parallel to the standard curves obtained using the kit standards. These results indicate that the immunoassay kit can be used to determine relative mass values for natural human PROINSULIN.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for PROINSULIN has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any PROINSULIN present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated antibody specific for PROINSULIN is added to the wells. Following a wash to remove any unbound antibody-biotin reagent, HRP link Streptavidin is added to the wells. After washing away any unbound enzyme, a substrate solution is added to the wells and color develops in proportion to the amount of PROINSULIN bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

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_ The kit should not be used beyond the expiration date on the kit label.

_ Do not mix or substitute reagents with those from other lots or sources.

_ It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.

_ If samples generate values higher than the highest standard, dilute the samples with Dilution Buffer and repeat the assay.

_ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

_ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors

have been tested in the immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
PROINSULIN Microplate - 96 well polystyrene microplate (12 strips of 8 wells) coated with an antibody against PROINSULIN.	095-01-01	1 plate
PROINSULIN Standard – 15 ng/vial of recombinant human PROINSULIN in a buffered protein base with preservative; lyophilized.	095-01-02	1 vial
Detection Antibody Concentrate – 1.05 mL/vial, 10-fold concentrate of biotinylated antibody against PROINSULIN with preservative; lyophilized.	095-01-03	1 vial
Positive Control - one vial of recombinant human PROINSULIN; lyophilized.	095-01-04	1 vial
Streptavidin-HRP Conjugate - 60 µL/vial, 200-fold concentrated solution of Streptavidin conjugate to HRP.	SAHRP	1 vial
Dilution Buffer – 60 mL of buffered protein based solution with preservative.	DB01	1 bottle
Wash Buffer - 50 mL of 10-fold concentrated buffered surfactant, with preservative.	WB01	1 bottle
TMB Substrate Solution - 11 mL of TMB substrate solution.	TMB01	1 bottle
Stop Solution - 11 mL of 0.5M HCl solution.	S-STOP	1 bottle
Plate Sealer	EAPS	1 piece
Plastic Pouch	P01	1 piece

STORAGE

Unopened Kit: Store at 2 – 8 °C for up to 8 months. For longer storage, unopened Standard, Positive Control and Detection Antibody Concentrate should be stored at -20 °C or -70 °C. Do not use kit past expiration date.

Opened / Reconstituted Reagents: Reconstituted Standard (Stock) and Detection Antibody concentrated solution SHOULD BE STORED at -20 °C or -70 °C for up to one month. Streptavidin-HRP Conjugate 200-fold concentrated solution (**protect from light**) and other components may be stored at 2 – 8 °C for up to 8 months.

Microplate Wells: Return unused wells to the plastic pouch with the desiccant pack. Microplate may be stored for up to 6 months at 2 – 8 °C after opening.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at ≤-20° C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20° C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤-20° C. Avoid repeated freeze-thaw cycles.

Optional: Use Aprotinin (enzyme inhibitor) for ALL sample collection to prevent sample degradation. 0.5 TIU per ml of sample solution.

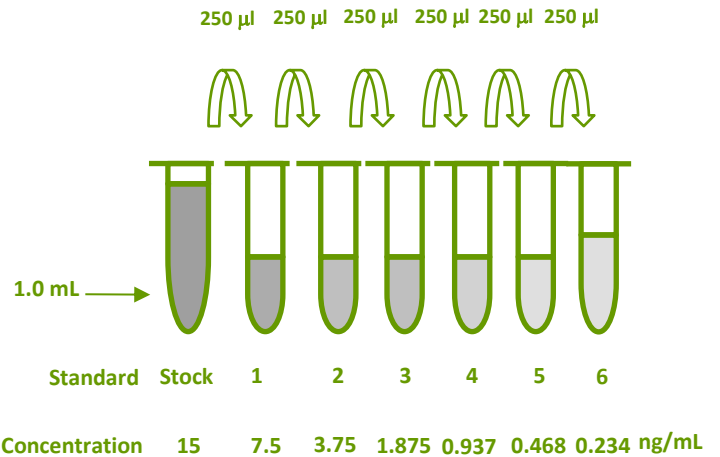
REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of 1x Wash Buffer.

PROINSULIN Standard - Refer to vial label for reconstitution volume. Reconstitute the **PROINSULIN** standard with 1.0 mL of Dilution Buffer. This reconstitution produces a stock solution of 15 ng/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 µL of the appropriate Dilution Buffer into tubes #1 to #6. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The **15 ng/mL** standard serves as the high standard. The appropriate Dilution Buffer serves as the zero standard (0 ng/mL).

TUBE	STANDARD	DILUTION BUFFER	CONCENTRATION
stock	powder	1.0 ml	15 ng/ml
# 1	250µl of stock	250µl	7.5 ng/ml
# 2	250µl of 1	250µl	3.75 ng/ml
# 3	250µl of 2	250µl	1.875 ng/ml
# 4	250µl of 3	250µl	0.9375 ng/ml
# 5	250µl of 4	250µl	0.468 ng/ml
# 6	250µl of 5	250µl	0.234 ng/ml



Positive Control - Reconstitute the **Positive Control** with 1.0 mL of Dilution Buffer. **Note:** Positive Control could be reused within a few days if stored at -20 °C ~ -70 °C.

Detection Antibody - Reconstitute the **Detection Antibody Concentrate** with 1.05 mL of Dilution Buffer to produce a 10-fold concentrated stock solution. Pipette 9.45 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 1.05 mL of 10-fold

concentrated stock solution to prepare working solution.

Streptavidin-HRP Conjugate - Pipette 11.94 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 60 µL of 200-fold concentrated stock solution to prepare working solution. **Note:** 1x working solution of Streptavidin-HRP Conjugate should be used within a few days (**protect from light**).

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards, positive control and samples be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the plastic pouch with the desiccant pack.
3. Add 100 µL of Dilution Buffer to Blank wells (F4, F5).
4. Add 100 µL of each standard solutions in reverse order of serial dilution (B2, B3 to G2, G3 and G4, G5), sample, or positive control (E4, E5) per well. Cover with plate sealer. Incubate for 2 hours on microplate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with 1x Wash Buffer (300 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100 µL of Detection Antibody working solution to each well. Cover with plate sealer. Incubate for 2 hours on microplate shaker at room temperature.
7. Repeat the aspiration/wash as in step 5.
8. Add 100 µL of **Streptavidin-HRP Conjugate working solution** to each well. Incubate for 60 minutes on microplate shaker at room temperature. **Protect from light.**
9. Repeat the aspiration/wash as in step 5.
10. Add 100 µL of Substrate Solution to each well. Incubate for 1-4 minutes on microplate shaker at

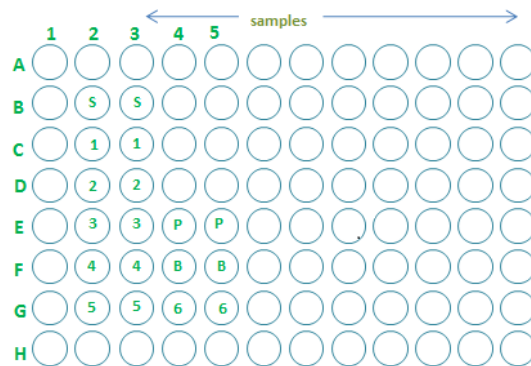
room temperature. **Protect from light. *Be ready to stop reaction with Stop Solution due to fast color development.**

11. Add 100 µL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
12. Determine the optical density of each well within 15 minutes, using a microplate reader set to 450 nm.

CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control and sample, and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the PROINSULIN concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.



CALIBRATION

This immunoassay is calibrated against a highly purified recombinant human PROINSULIN.

SENSITIVITY

The minimum detectable dose (MDD) of PROINSULIN was 80 pg/mL.

TYPICAL DATA

This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

STANDARD (NG/ML)	AVERAGE OD450 (CORRECTED)
Blank	0 (0.089)
0.234	0.035
0.468	0.064
0.9375	0.127
1.875	0.249
3.75	0.476
7.5	0.934
15	1.863

SPECIFICITY

PROTEINS	CROSS-REACTIVITY
Human Proinsulin	100%
Human insulin	0
Human IGF-I	0
Human IGF-II	0
Human relaxin	0

SUMMARY OF ASSAY PROCEDURE

