

HUMAN CYSTATIN C ELISA KIT

FOR THE QUANTITATIVE DETERMINATION
OF HUMAN CYSTATIN C CONCENTRATIONS
IN CELL CULTURE SUPERNATES, EDTA
PLASMA, SERUM AND URINE



ALWAYS REFER TO LOT SPECIFIC PROTOCOL
PROVIDED WITH EACH KIT FOR
INSTRUCTIONS. PROTOCOL MUST BE
READ BEFORE USING THIS PRODUCT.

FOR RESEARCH USE ONLY. NOT FOR USE IN
DIAGNOSTIC PROCEDURES.

PRODUCT INFORMATION:

| | |
|---|---|
| ELISA NAME | HUMAN CYSTATIN C ELISA |
| Catalog No. | SK00699-01 |
| Lot No. | 20111927 |
| Formulation | 96T |
| Standard range | 15.6 - 1000 pg/mL |
| Sensitivity | 15.6 pg/mL |
| Sample Volume | 100 µL |
| Sample Type | Cell Culture Supernates, EDTA Plasma, Serum and Urine |
| Dilution Factor | Serum, EDTA Plasma: 4000 - 8000 Urine: Pretest (Optimal dilutions should be determined by each laboratory for each application) |
| Specificity | Human Cystatin C |
| Calibration | Human Cystatin C |
| Intra-assay Precision | 4 - 6% |
| Inter-assay Precision | 8 - 12% |
| Storage | 2 – 8° C |
| This kit contains sufficient materials to run 35 samples duplicated provided that assay is run according to protocol. | |

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DESCRIPTION

This Human Cystatin C ELISA Kit contains the necessary components required for the quantitative measurement of recombinant and/or natural human Cystatin C from cell culture supernates, serum, EDTA plasma and urine in a sandwich ELISA format.

This immunoassay contains human Cystatin C and antibodies raised against this protein. Results from this immunoassay have shown to accurately quantify recombinant and natural Cystatin C samples.

ASSAY OVERVIEW

This assay employs the quantitative sandwich ELISA format. The plate is pre-coated with an antibody specific for human Cystatin C. The capture antibody can bind to the human Cystatin C in the standard and samples. After washing the plate of any unbound substances, a biotinylated antibody against Cystatin C is added to the wells. After another washing of the plate, Streptavidin-HRP Conjugate is added. After the last wash to remove any unbound enzyme, a substrate solution is added to the wells and color develops in direct proportion to the amount of human Cystatin C bound in the standard solutions or samples. A standard curve can be established and sample values can be read off the standard curve.

PROCEDURAL LIMITATIONS

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_This ELISA kit should not be used beyond the expiration date on the kit label.

_Do not mix reagents with those from other lots or sources.

_It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.

_Each laboratory must determine the optimal dilution factors for the samples being assayed with a pretest. If samples generate values that are not within the dynamic range of the standard curve, further concentrate or dilute the samples as required with Dilution Buffer and repeat the assay.

_Any modifications in buffers, pipetting technique, washing technique, incubation time or temperature, as well as kit age can cause a change in signal.

_Not all interfering factors have been tested in the immunoassay, therefore the possibility of interference cannot be excluded.

COMPONENTS PROVIDED

| DESCRIPTION | CODE | QUANTITY |
|--|------------------|------------------|
| Cystatin C Microplate - 96 well polystyrene microplate (12 strips of 8 wells) coated with an antibody against Cystatin C. | 699-01-01 | 1 plate |
| Cystatin C Standard – 4000 pg/vial of human Cystatin C in a buffered protein base with preservative; lyophilized. | 699-01-02 | 1 vial |
| Detection Antibody Concentrate – 1.2 mL/vial, 10-fold concentrate of biotinylated antibody against Cystatin C with preservative; lyophilized. | 699-01-03 | 1 vial |
| Positive Control - one vial of human Cystatin C; lyophilized. | 699-01-04 | 1 vial |
| Streptavidin-HRP Conjugate - 120 µl/vial, 100-fold concentrated solution of Streptavidin conjugate to HRP. | SAHRP | 1 vial |
| Dilution Buffer – 60 mL of buffered protein based solution with preservative. | DB10 | 2 bottles |
| HRP Diluent Solution – 12 mL of buffered protein based solution with preservative. | DB08 | 1 bottle |
| Wash Buffer - 50 mL of 10-fold concentrated buffered surfactant, with preservative. | WB01 | 1 bottle |
| TMB Substrate Solution - 11 mL of TMB substrate solution. | TMB01 | 1 bottle |
| Stop Solution - 11 mL of 0.5M HCl. | S-STOP | 1 bottle |
| Plate Sealer | EAPS | 1 |
| Plastic Pouch | P01 | 1 |

STORAGE

Unopened Kit: Store at 2 – 8° C for up to 6 months. For longer storage, unopened Standard, Positive Control and Detection Antibody Concentrate should be stored at -20° C or -70° C. Do not use kit past expiration date.

Opened / Reconstituted Reagents: Reconstituted Standard (stock) solution and Detection Antibody concentrated solution SHOULD BE STORED at -20° C

or -70° C for up to one month. SAHRP Conjugate 100-fold concentrated solution and Substrate Solution can be stored at 2 – 8° C for up to 6 months (**DO NOT FREEZE** and **PROTECT FROM LIGHT**). All other components may be stored at 2 – 8° C for up to 6 months.

Microplate Wells: Return unused strips to the plastic pouch with the desiccant pack. Microplate may be stored for up to 6 months at 2 – 8° C.

ADDITIONAL MATERIALS REQUIRED

- Microplate reader capable of absorbance measurement at 450 nm.
- Microplate shaker (250 – 300 rpm).
- Microplate washer or manifold dispenser.
- 100 mL and 500 mL graduated cylinders.
- Multi-channel Pipette, Pipettes and pipette tips.
- Deionized or distilled water.

PRECAUTION

This kit should be handled by those persons who have been trained in and can follow the principles of good laboratory practice. Wear protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken while handling solutions in this kit to avoid contact with skin or eyes, especially with the stop solution because it contains diluted hydrochloric acid. Wash immediately with water in case of contact on skin or eyes.

SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at $\leq -20^{\circ}$ C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}$ C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at $\leq -20^{\circ}$ C. Avoid repeated freeze-thaw cycles.

Urine - Freshly collected urine samples were allowed to sit at room temperature for 30 minutes to sediment, and the supernatant was aliquoted and stored at -70° C until analysis. Centrifuge again before assaying to remove any additional precipitates that may appear after storage.

Optional: Use Aprotinin (enzyme inhibitor) for ALL sample collection to prevent sample degradation. 0.5 TIU per ml of sample solution.

SAMPLE PREPARATION

Serum and plasma samples may require a 4000 ~ 8000-fold dilution. A suggested 10-fold dilution is 10 μ L sample + 90 μ L Dilution Buffer; then, 10 μ L of 10-fold diluted sample + 90 μ L Dilution Buffer to make 100-fold diluted sample. Lastly, 6 μ L of 100-fold diluted sample + 234 μ L Dilution Buffer to make 4000-fold diluted sample. To make an 8000-fold dilution is 3 μ L of 100-fold diluted sample + 237 μ L Dilution Buffer.

Urine samples will require a pretest to determine the best dilution.

Optimal dilutions should be determined by each laboratory for each application.

Use polypropylene test tubes.

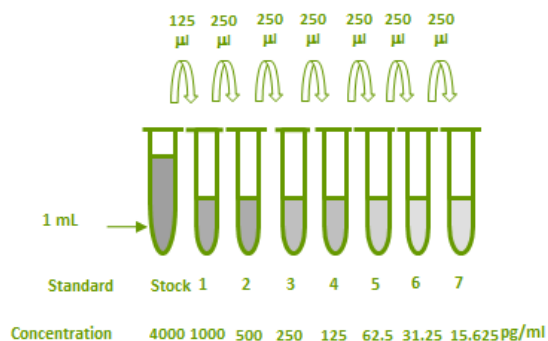
REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of 1x Wash Buffer.

Cystatin C Standard - Reconstitute the Cystatin C standard with 1.0 mL of Dilution Buffer. This reconstitution produces a stock solution of 4000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 375 μ L of Dilution Buffer into tube #1, then transfer 125 μ L of 4000 pg/mL stock solution to make the high standard of 1000 pg/mL. Pipette 250 μ L of Dilution Buffer into tubes #2 to #7. Use the high standard of 1000 pg/mL to produce a dilution series (next page). Mix each tube thoroughly before the next transfer. The **1000 pg/mL** standard serves as the high standard. The Dilution Buffer serves as the zero standard (0 pg/mL).

| TUBE | STANDARD | DILUTION BUFFER | CONCENTRATION |
|-------|-----------------|-----------------|---------------|
| stock | Powder | 1000 µl | 4000 pg/ml |
| # 1 | 125 µl of stock | 375 µl | 1000 pg/ml |
| # 2 | 250 µl of 1 | 250 µl | 500 pg/ml |
| # 3 | 250 µl of 2 | 250 µl | 250 pg/ml |
| # 4 | 250 µl of 3 | 250 µl | 125 pg/ml |
| # 5 | 250 µl of 4 | 250 µl | 62.5 pg/ml |
| # 6 | 250 µl of 5 | 250 µl | 31.25 pg/ml |
| # 7 | 250 µl of 6 | 250 µl | 15.625 pg/ml |



Positive Control - Reconstitute the Positive Control with 1.0 mL of Dilution Buffer. **Note:** Positive Control could be reused within a few days if stored at -20° C or -70° C.

Detection Antibody Concentrate - Reconstitute the Detection Antibody Concentrate with 1.2 mL of Dilution Buffer to produce a 10-fold concentrated stock solution. Pipette 9.45 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 1.05 mL of 10-fold concentrated stock solution to prepare working solution.

Streptavidin-HRP Conjugate - Pipette 11.88 mL of **HRP Diluent Solution (DB08)** into a 15 mL centrifuge tube and transfer 120 µL of 100-fold concentrated stock solution to prepare working solution. **Note:** 1x working solution of Streptavidin-HRP Conjugate should be used within a few days. **(Protect from light)**

ELISA PROTOCOL

Bring all reagents and samples to room temperature before the start of the assay. Blank, standard dilutions, positive control and samples should be assayed in duplicate. ELISA Protocol may need further optimization.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the plastic pouch with the desiccant pack.
3. Add 100 µL per well of Dilution Buffer to Blank wells.
4. Add 100 µL of Standard dilutions, samples, or positive control per well. Cover with plate sealer. Incubate for 2 hours on microplate shaker at room temperature.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with 1x Wash Buffer (300 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100 µL of Detection Antibody working solution to each well. Cover with plate sealer. Incubate for 2 hours on microplate shaker at room temperature.
7. Repeat the aspiration/wash as in step 5.
8. Add 100 µL of Streptavidin-HRP Conjugate working solution to each well. Incubate for 45 minutes on microplate shaker at room temperature. **Protect from light.**
9. Repeat the aspiration/wash as in step 5.
10. Add 100 µL of Substrate Solution to each well. Incubate for 2-6 minutes on microplate shaker at room temperature. **Protect from light.**
11. Add 100 µL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
12. Determine the optical density of each well within 15 minutes, using a microplate reader set to 450 nm.

CALCULATION OF RESULTS

Create a standard curve by plotting the log of the known concentrations of the standard dilutions (x-axis) versus the log of its corresponding O.D. (y-axis) and draw the best fit line through the points. It is recommended to use computer software capable of generating a log-log curve fit to more accurately quantify the standard dilutions.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

TYPICAL DATA

This standard curve is provided for demonstration only. A new standard curve should be generated for each set of samples assayed.

| CYSTATIN C (PG/ML) | AVERAGE OD450 (CORRECTED)* |
|--------------------|----------------------------|
| Blank | 0 (0.062) |
| 15.625 | 0.107 |
| 31.25 | 0.232 |
| 62.5 | 0.444 |
| 125 | 0.783 |
| 250 | 1.385 |
| 500 | 2.334 |
| 1000 | 3.162 |

SPECIFICITY

| PROTEINS | CROSS-REACTIVITY (%) |
|-----------------------|----------------------|
| Human Cystatin C | 100 |
| Mouse Cystatin C | 0 |
| Human Cathepsin B | 0 |
| Human Cathepsin X/Z/P | 0 |
| Human Cystatin A | 0 |
| Human Cystatin B | 0 |
| Human Cystatin D | 0 |
| Human Cystatin E/M | 0 |

SUMMARY OF ASSAY PROCEDURE

