

## HUMAN CCL18/PARC ELISA KIT

FOR THE QUANTITATIVE DETERMINATION  
OF HUMAN CCL18/PARC  
CONCENTRATIONS IN SERUM AND EDTA  
PLASMA



### PURCHASE INFORMATION:

ELISA Name	Human CCL18/PARC ELISA
Catalog No.	SK00466-01
Lot No.	
Formulation	96 T
Standard range	15.6-1000 pg/ml
Sensitivity	5 pg/mL
Sample Volume	100 µl
Sample Type	Serum, EDTA Plasma
Dilution Factors	200 (Optimal dilutions should be determined by each laboratory for each application)
Specificity	Human CCL18 only
Intra-assay Precision	4 - 6%
Inter-assay Precision	8 - 10%
Storage	2°C - 8°C

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DIAGNOSTIC PROCEDURES.

## INTRODUCTION

Human CCL18/PARC immunoassay is a 3.5 - 4.5 hour solid phase ELISA designed to measure human CCL18/PARC in serum, and EDTA plasma. It contains recombinant human CCL18/PARC and antibodies raised against this protein. It has been shown to accurately quantify recombinant human CCL18/PARC. Results obtained with naturally occurring CCL18/PARC samples showed linear curves that were parallel to the standard curves obtained using the kit standards. These results indicate that the immunoassay kit can be used to determine relative mass values for natural human CCL18/PARC.

## PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for CCL18/PARC has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any CCL18/PARC present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated antibody specific for CCL18/PARC is added to the wells. Following a wash to remove any unbound antibody-biotin reagent, HRP link Streptavidin is added to the wells. After washing away any unbound enzyme, a substrate solution is added to the wells and color develops in proportion to the amount of CCL18/PARC bound in the initial step. The color development is stopped and the intensity of the color is measured.

## LIMITATIONS OF THE PROCEDURE

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\_ The kit should not be used beyond the expiration date on the kit label.

\_ Do not mix or substitute reagents with those from other lots or sources.

\_ It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.

\_ If samples generate values higher than the highest standard, dilute the samples with Dilution Buffer and repeat the assay.

\_ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

\_ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors

have been tested in the immunoassay, the possibility of interference cannot be excluded.

## MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
<b>CCL18/PARC Microplate</b> - 96 well polystyrene microplate (12 strips of 8 wells) coated with a purified antibody against human CCL18/PARC.	<b>466-01-01</b>	<b>1 plate</b>
<b>CCL18/PARC Standard</b> - 1ng/vial of recombinant human CCL18/PARC in a buffered protein base with preservatives; lyophilized.	<b>466-01-02</b>	<b>2vials</b>
<b>Detection Antibody Concentrate</b> - 1.05 mL/vial, 10-fold concentrate of biotinylated antibody against human CCL18/PARC with preservatives; lyophilized.	<b>466-01-03</b>	<b>1 vial</b>
<b>Positive Control</b> - one vial of recombinant human CCL18/PARC, lyophilized	<b>466-01-04</b>	<b>1 vial</b>
<b>Streptavidin-HRP Conjugate</b> - 60 µL/vial, 200-fold concentrated solution of Streptavidin conjugate to HRP with preservatives	<b>SAHRP</b>	<b>1 vial</b>
<b>Dilution Buffer</b> - 60 mL of buffered protein based solution with preservatives	<b>DB01</b>	<b>1 bottle</b>
<b>Wash Buffer</b> - 50 mL of 10-fold concentrated buffered surfactant, with preservative.	<b>WB01</b>	<b>1 bottle</b>
<b>TMB Substrate Solution</b> - 11 mL of TMB substrate solution	<b>TMB01</b>	<b>1 bottle</b>
<b>Stop Solution</b> - 11 mL of 0.5M HCl	<b>S-STOP</b>	<b>1 bottle</b>
<b>Plate Sealer</b>	<b>EAPS</b>	<b>1</b>
<b>Plastic Pouch</b>	<b>P01</b>	<b>1</b>

## STORAGE

**Unopened Kit:** Store at 2 - 8° C for up to 8 months. For longer storage, unopened Standard, Positive Control and Detection Antibody Concentrate should be stored at -20°C or -70°C. Do not use kit past expiration date.

**Opened / Reconstituted Reagents:** Reconstituted Standard and Detection Antibody Concentrate

Solution SHOULD BE STORED at -20°C or -70°C for up to one month. Streptavidin-HRP Conjugate 200-fold Concentrate (**protect from light**) and other components may be stored at 2 - 8°C for up to 8 months.

**Microplate Wells:** Return unused wells to the plastic pouch with the desiccant pack and seal along entire edge of zip-seal. Microplate may be stored for up to 6 months at 2 - 8°C after opening.

### OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

### PRECAUTIONS FOR USE

All reagents should be considered as potentially hazardous. The stop solution contains diluted hydrochloric acid. Appropriate care should be taken while handling this solution. We recommend that this product be handled by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water.

### SAMPLE COLLECTION AND STORAGE

**Serum** - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20°C. Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20°. Avoid repeated freeze-thaw cycles.

**Note: Use Aprotinin (enzyme inhibitor) (Code No.: 00700-01-25) for ALL sample collection to prevent**

**sample degradation. 0.5 TIU per ml of sample solution.**

### SAMPLE PREPARATION

Serum and plasma samples require a 200-fold dilution. A suggested 200-fold dilution is 10 µL sample + 190 µL Dilution Buffer. Following 30 µL of 20-fold diluted sample solution + 270 µL Dilution Buffer.

**Optimal dilutions should be determined by each laboratory for each application.**

**Use polypropylene test tubes.**

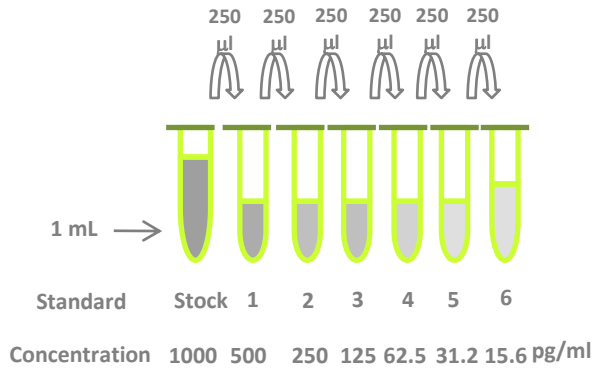
### REAGENT PREPARATION

**Bring all reagents to room temperature before use.**

**Wash Buffer** - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of Wash Buffer.

**CCL18/PARC Standard - Refer to vial label for reconstitution volume.** Reconstitute the **CCL18/PARC Standard** with 1 mL of Dilution Buffer. This reconstitution produces a stock solution of 1000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 µL of Dilution Buffer into tubes #1 to #7. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 1000 pg/mL standard serves as the high standard. The Dilution Buffer serves as the zero standard (0 pg/mL).

Tube	Standard	Dilution Buffer	Concentration
stock	Powder	1000 µl	1000 pg/ml
# 1	250 µl of stock	250 µl	500 pg/ml
# 2	250 µl of 1	250 µl	250 pg/ml
# 3	250 µl of 2	250 µl	125 pg/ml
# 4	250 µl of 3	250 µl	62.5 pg/ml
# 5	250 µl of 4	250 µl	31.25 pg/ml
# 6	250 µl of 5	250 µl	15.6 pg/ml



**Detection Antibody** - Reconstitute the Detection Antibody Concentrate with 1.05 mL of Dilution Buffer to produce a 10-fold concentrated stock solution. Pipette 9.45 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 1.05 mL of 10-fold concentrated stock solution to prepare working solution.

**Streptavidin-HRP Conjugate** - Pipette 11.94 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 60 µL of 200-fold concentrated stock solution to prepare working solution. **Note:** 1x working solution of Streptavidin-HRP Conjugate should be used within a few days (**protect from light**).

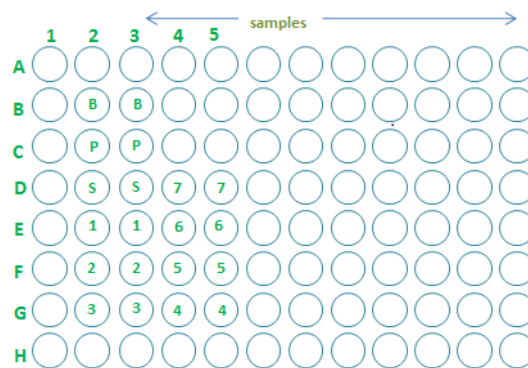
**Positive Control** - Reconstitute the Positive Control with 1 mL of Dilution Buffer. **Note:** Positive Control should be prepared and used immediately.

**ASSAY PROCEDURE**

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards, positive control and samples be assayed in duplicates.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess micro-plate strips from the plate frame, return them to the plastic pouch with the desiccant pack and seal.
3. Add 100 µL of Dilution Buffer to Blank wells (B2, B3).
4. Add 100 µL of Standard (D2, D3 to G2, G3 and G4, G5 to D4, D5), sample, or positive control (C2, C3) per well. Cover with plate sealer. Incubate for 2 hours on micro-plate shaker at room temperature. A plate layout is provided to record standards and samples assayed.

5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (300 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100 µL of Detection Antibody working solution to each well. Cover with plate sealer. Incubate for 2 hours on micro-plate shaker at room temperature.
7. Repeat the aspiration/wash as in step 5.
8. Add 100 µL of Streptavidin-HRP Conjugate working solution to each well. Incubate it on microplate shaker for 60 minutes at room temperature. **Protect from light.**
9. Repeat the aspiration/wash as in step 5.
10. Add 100 µL of Substrate Solution to each well. Incubate for 12-18 minutes on microplate shaker at room temperature. **Protect from light.**
11. Add 100 µL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
12. Determine the optical density of each well within 15 minutes, using a micro-plate reader set to 450 nm.



**CALCULATION OF RESULTS**

Average the duplicate readings for each standard, positive control and samples, and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-

axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the CCL18/PARC concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted BY 200, the concentration read from the standard curve must be multiplied by the dilution factor 200.

### TYPICAL DATA

This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

STANDARD (PG/ML)	AVERAGE OD450 (CORRECTED)
Blank	0(0.097)
15.6	0.040
31.25	0.084
62.5	0.174
125	0.368
250	0.701
500	1.347
1000	2.631

- Lot No.
- Positive Control:

### CALIBRATION

This immunoassay is calibrated against a highly purified recombinant human CCL18/PARC.

### SENSITIVITY

Twenty-five assays were evaluated and the minimum detectable dose (MDD) of CCL18/PARC was 5 pg/mL.

### SPECIFICITY

This assay recognizes both natural and recombinant human CCL18/PARC. No significant cross-reactivity or interference was observed.

Proteins	Cross-reactivity (%)
Human CCL18/PARC	100
Human MCP-1	0
Human IL-6	0

### SUMMARY OF ASSAY PROCEDURE

PREPARE REAGENTS, SAMPLES AND STANDARDS
↓
Add 100 µl of <b>standard, samples, positive control</b> to each well. Incubate 2 hours on the plate shaker at RT.
↓
Aspirate and wash 4 times.
↓
Add 100 µl <b>Detection Antibody working solution</b> to each well. Incubate 2 hours on the plate shaker at RT.
↓
Aspirate and wash 4 times.
↓
Add 100 µl <b>Streptavidin-HRP conjugate working solution</b> to each well. Incubate 60 min on the plate shaker at RT. <b>Protect from light.</b>
↓
Aspirate and wash 4 times.
↓
Add 100 µl <b>Substrate Solution</b> to each well. Incubate 12-18 mins on the plate shaker. <b>Protect from light.</b>
↓
Add 100 µl <b>Stop Solution</b> to each well. Read 450nm within 15 min

### REFERENCE

1. Lota HK, Renzoni EA. Circulating biomarkers of interstitial lung disease in systemic sclerosis. *Int J Rheumatol.* 2012;2012:121439. Epub 2012 Sep 3.
2. de Jager SC, ET AL. Chemokines CCL3/MIP1 $\alpha$ , CCL5/RANTES and CCL18/PARC are Independent Risk Predictors of Short-Term Mortality in Patients with Acute Coronary Syndromes. *PLoS One.* 2012;7(9):e45804. doi: 10.1371/journal.pone.0045804. Epub 2012 Sep 21.
3. Plönes T Serum level of CC-chemokine ligand 18 is increased in patients with non-small-cell lung cancer and correlates with survival time in adenocarcinomas. *PLoS One.* 2012;7(7):e41746. Epub 2012 Jul 25.