

MOUSE SOLUBLE CD14 ELISA KIT

FOR THE QUANTITATIVE DETERMINATION OF MOUSE SOLUBLE CD14 CONCENTRATIONS IN CELL CULTURES, SERUM AND PLASMA.



FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

PURCHASE INFORMATION:

ELISA NAME	MOUSE SOLUBLE CD14 ELISA KIT
Catalog No.	SK00178-03
Lot No.	
Formulation	96 T
Standard range	78-5000 pg/mL
Sensitivity	20 pg/mL
Sample Volume	100 µl
Sample Type	Serum, Plasma, Cell Culture Supernates
Dilution factor	Optimal dilutions should be determined by each laboratory for each application
Specificity	Mouse sCD14
Intra-assay Precision	4-6%
Inter-assay Precision	8-10%
Storage	2 - 8 °C

ORDER CONTACT:
 AVISCERA BIOSCIENCE INC.
 2348 Walsh Ave., Suite C
 Santa Clara, CA 95051
 Tel: (408) 982 0300
 Fax: (408) 982 0301
 Email: Sales@AvisceraBioscience.com
 Info@AvisceraBioscience.com
www.AvisceraBioscience.com

INTRODUCTION

Mouse Soluble CD14 immunoassay is a 3.5 - 4.5 hour solid phase ELISA designed to measure mouse sCD14 in cell culture supernates, serum and EDTA plasma. It contains recombinant mouse sCD14 and antibodies raised against this protein. It has been shown to accurately quantify recombinant mouse sCD14. Results obtained with naturally occurring mouse sCD14 samples showed linear curves that were parallel to the standard curves obtained using the kit standards. These results indicate that the immunoassay kit can be used to determine relative mass values for natural mouse sCD14.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for mouse sCD14 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any sCD14 present is bound by the immobilized antibody. After washing away any unbound substances, an antibody HRP conjugate specific for mouse sCD14 is added to the wells. After washing away any unbound enzyme, a substrate solution is added to the wells and color develops in proportion to the amount of mouse sCD14 bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

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_The kit should not be used beyond the expiration date on the kit label.

_ Do not mix or substitute reagents with those from other lots or sources.

_It is important that the DILUTION BUFFER selected for the standard curve be consistent with the samples being assayed.

_If samples generate values higher than the highest standard, dilute the samples with the appropriate DILUTION BUFFER and repeat the assay.

_Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

_This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors have been tested in the Immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
Mouse sCD14 Microplate - 96 well polystyrene microplate (12 strips of 8 wells) coated with antibody against mouse sCD14.	178-03-01	1 plate
Mouse Standard Standard – 5 ng/vial of recombinant mouse sCD14 in a buffered protein base with preservatives; lyophilized.	178-03-02	1 vial
Detection Antibody Concentrate – 105 µL / vial, 100-fold concentrated of antibody HRP conjugate against mouse sCD14 with preservatives;	178-03-03	1 vial
Positive Control - one vial of recombinant mouse sCD14 with preservatives; lyophilized.	178-03-04	1 vial
Dilution Buffer - 60 mL of buffered protein based solution with preservatives	DB10	1 bottle
Wash Buffer - 50 mL of 10-fold concentrated buffered surfactant, with preservative.	WB01	1 bottle
TMB Substrate Solution - 11 mL of TMB substrate solution	TMB01	1 bottle
Stop Solution - 11 mL of 0.5M HCl	S-STOP	1 bottle
Plate sealer	EAPS	1 piece
Plastic Pouch	P01	1 piece

STORAGE

Unopened Kit: Store at 2 - 8° C for up to 12 months. For longer storage, unopened Standard, Positive control and Detection Antibody Concentrated should be stored at -20 or -70 °C. Do not use past kit expiration date.

Opened / Reconstituted Reagents: Reconstituted Standard, Detection Antibody Concentrate Solution SHOULD BE STORED at -20 °C or – 70°C for up to one month.

The detection antibody - HRP Conjugate 100-fold concentrated and other components may be stored at 2 - 8°C for up to 12 months.

Microplate Wells: Return unused wells to the plastic pouch with the desiccant pack, reseal along the entire edge of zip-seal. Microplate may be stored for up to 6 months at 2 - 8° C after opening.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

PRECAUTIONS FOR USE

All reagents should be considered as potentially hazardous. The stop solution contains diluted Hydrochloric acid. Appropriate care, therefore, should be taken while handling this solution. We therefore recommend that this product is handled only by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water.

SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at $\leq -20^{\circ}$ C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}$ C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at $\leq -20^{\circ}$ C. Avoid repeated freeze-thaw cycles.

Note: Use Aprotinin (enzyme inhibitor) for ALL sample collection to prevent sample degradation. 0.5 TIU per ml of sample solution.

SAMPLE PREPARATION

Optimal dilutions should be determined by each laboratory for each application. Use polypropylene test tubes

REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of Wash Buffer.

Mouse sCD14 Standard - Refer to vial label for reconstitution volume. Reconstitute the Mouse sCD14 Standard with 1 mL of Dilution Buffer. This reconstitution produces a stock solution of 5000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 μ L of the appropriate Sample Solution into the tube #1 to #6. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 5000 pg/mL standard serves as the high standard. The appropriate Sample Solution serves as the zero standard (0 pg/mL).

TUBE	STANDARD	DILUTION BUFFER	CONCENTRATION
Stock	powder	1000 μ l	5000 pg/ml
# 1	250 μ l of stock	250 μ l	2500 pg/ml
# 2	250 μ l of 1	250 μ l	1250 pg/ml
# 3	250 μ l of 2	250 μ l	625 pg/ml
# 4	250 μ l of 3	250 μ l	312.5 pg/ml
# 5	250 μ l of 4	250 μ l	156 pg/ml
# 6	250 μ l of 5	250 μ l	78 pg/ml

Detection Antibody HRP conjugate Concentrate -

Pipette 10.395 mL of Dilution Buffer into the 15 ml centrifuge tube and transfer 105 µl of 100-fold concentrated stock solution to prepare working solution.

Positive Control - Reconstitute the positive control with 1mL of Dilution Buffer to make positive control solution.

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards, positive control and samples be assayed in duplicates.

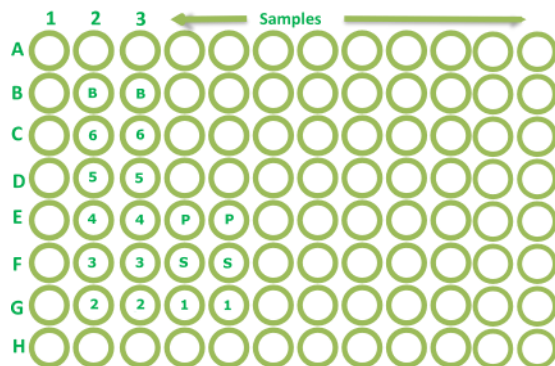
1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess micro-plate strips from the plate frame, return them to the plastic pouch with the desiccant pack, reseal.
3. Add 100 µL of Dilution Buffer to Blank well (B2, B3).
4. Add 100 µL of Standard (from C2, C3 to G2, G3, F4, F5 to G4, G5), samples, or positive control per well (E4, E5). Cover with plate sealer. Incubate for 2 hours on a micro-plate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Buffer (300 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100 µL of Detection Antibody HRP conjugate working solution to each well. Cover with sealer. Incubate for 1 hour on micro-plate shaker at room temperature. **Protect from light.**
7. Repeat the aspiration/wash as in step 5.
8. Add 100 µL of Substrate Solution to each well. Incubate for 12-18 minutes at room temperature. **Protect from light.**
9. Add 100 µL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.

12. Determine the optical density of each well within 15 minutes, using a micro-plate reader set to 450 nm.

CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control, and sample and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the mouse sCD14 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.



TYPICAL DATA

This standard curve* is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

STD (PG/ML)	CORRECTED (450NM)
Blank	0 (0.094)
78	0.029
156	0.064
312.5	0.138
625	0.277
1250	0.502
2500	0.899
5000	1.684

Lot No.:

- Positive Control :

CALIBRATION

This immunoassay is calibrated against a highly purified recombinant mouse sCD14

SENSITIVITY

Twenty-five assays were evaluated and the minimum detectable dose (MDD) of mouse sCD14 was 20 pg/mL.

SPECIFICITY

PROTEINS	CROSSREACTIVITY (%)
Mouse sCD14	100
Human sCD14	0
Mouse sCD36	0
Mouse sRAGE	0

SUMMARY OF ASSAY PROCEDURE

PREPARE REAGENTS, SAMPLES AND STANDARDS
↓
Add 100 µl of standard, samples, positive control to each well. Incubate for 2 hours at RT.
↓
Aspirate and wash 4 times.
↓
Add 100 µl Detection Antibody HRP conjugate working solution to each well. Incubate for 1 hour on the plate shaker at RT. Protect from light.
↓
Aspirate and wash 4 times.
Add 100 µl Substrate Solution to each well. Incubate 12-18 min on the bench top. Protect from light.
↓
Add 100 µl Stop Solution to each well. Read 450nm within 15 min